

# ERRATA

Page Number	Title	Section	Description
<b>USP 34-NF 29</b>			
309	(741) <i>Melting Range or Temperature</i>		Line 26: Change "Five" to: Eight
407	(921) <i>Water Determination</i>	<i>Method 1a (Direct Titration)</i>	Line 22 under <i>Standardization of the Reagent</i> : Change "For sodium tartrate, quickly add 20 to 125 mg of sodium tartrate (C <sub>4</sub> H <sub>4</sub> Na <sub>2</sub> O <sub>6</sub> ·2H <sub>2</sub> O), accurately weighed by difference, and titrate to the endpoint." to: For sodium tartrate dihydrate, quickly add 20 to 125 mg of sodium tartrate dihydrate (C <sub>4</sub> H <sub>4</sub> Na <sub>2</sub> O <sub>6</sub> ·2H <sub>2</sub> O), accurately weighed by difference, and titrate to the endpoint.
954	<i>Tetrahydro-2-furancarboxylic Acid</i>		Line 3: Change "[NOTE—A suitable grade is available from www.sigma-aldrich.com, catalog number 345117.]" to: [NOTE—A suitable grade is available from www.sigma-aldrich.com, catalog number 341517.]
1125	<i>Echinacea purpurea Aerial Parts</i>	<i>Botanical characteristics</i>	Line 3 under <i>Leaf</i> : Change "abundant on the dorsal surface and fewer on the ventral surface" to: abundant on the ventral surface and fewer on the dorsal surface
1166	<i>Glucosamine Tablets</i>	<i>Assay</i>	Line 3 under <i>Assay preparation</i> : Change "80 mg" to: 312 mg
1167	<i>Glucosamine Sulfate Potassium Chloride</i>	<i>Assay</i>	Line 1 under <i>Assay preparation</i> : Change "187.5 mg" to: 263 mg
1167	<i>Glucosamine Sulfate Sodium Chloride</i>	<i>Assay</i>	Line 1 under <i>Assay preparation</i> : Change "187.5 mg" to: 250 mg
1323	<i>Oil- and Water-Soluble Vitamins with Minerals Tablets</i>	<b>STRENGTH</b> <i>Cholecalciferol or Ergocalciferol (Vitamin D), Method 3</i>	Line 6 under <i>Analysis</i> : Change "Result = (r <sub>U</sub> /r <sub>S</sub> ) × (C <sub>S</sub> /C <sub>U</sub> ) × F × 100" to: Result = (r <sub>U</sub> /r <sub>S</sub> ) × (C <sub>S</sub> /C <sub>U</sub> ) × 100 Line 16 under <i>Analysis</i> : Delete "F = correction factor to account for the average amount of previtamin D present in the <i>Sample solution</i> , 1.09"
1520	<i>Erythritol</i>	<i>USP Reference standards (11)</i>	Change "USP Erythritol RS" to: USP Erythritol RS <i>meso</i> -Erythritol, 1,2,3,4-butanetetrol. C <sub>4</sub> H <sub>10</sub> O <sub>4</sub> 122.12
1969	<i>Azithromycin for Injection</i>	<i>USP Reference standards (11)</i>	Line 12: Add "USP Desosaminylazithromycin RS"
1971	<i>Azithromycin for Oral Suspension</i>	<i>USP Reference standards (11)</i>	Line 1: Add "USP Azaerythromycin RS"
2488	<i>Desflurane</i>	<b>IMPURITIES</b> <i>Organic Impurities</i>	Line 12 under <i>Analysis</i> : Change "r <sub>U</sub> = peak response of the Desflurane used as the solvent r <sub>S</sub> = peak response of the <i>Standard solution</i> " to: r <sub>U</sub> = peak response of each impurity from the Desflurane used as the solvent r <sub>S</sub> = peak response of each impurity from the <i>Standard solution</i>

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2488	<i>Desflurane</i>	IMPURITIES <i>Organic Impurities</i>	<p>Line 21 under <i>Analysis</i>: Change “<math>r_U</math> = peak response of the <i>Sample solution</i>”  <math>r_S</math> = peak response of the <i>Standard solution</i>  <math>C_F</math> = final concentration in the <i>Standard solution</i> (%)”  to:  <math>r_U</math> = peak response of each impurity from the <i>Sample solution</i>  <math>r_S</math> = peak response of each impurity from the <i>Standard solution</i>  <math>C_F</math> = final concentration of each impurity in the <i>Standard solution</i> (%)</p> <p>Line 26 under <i>Analysis</i>: Change “<math>r_U</math> = peak response of the <i>Sample solution</i>”  <math>r_S</math> = peak response of the <i>Standard solution</i>”  to:  <math>r_U</math> = peak response of each impurity from the <i>Sample solution</i>  <math>r_S</math> = peak response of Isoflurane from the <i>Standard solution</i></p>
2610	<i>Divalproex Sodium</i>	<i>USP Reference standards</i> (11)	<p>Line 2: Change “USP Divalproex Sodium RS  Pentanoic acid, 2-propyl-, sodium salt (2 : 1).  <math>(C_{16}H_{31}NaO_4)_n</math> Repeating unit molecular weight, 310.41”  to:  USP Divalproex Sodium RS  Sodium hydrogen bis(2-propylvalerate), oligomer; pentanoic acid,  2-propyl-, sodium salt (2 : 1).  <math>(C_{16}H_{31}NaO_4)_n</math> 310.41</p>
2782	<i>Etidronate Disodium</i>	<i>USP Reference standards</i> (11)	<p>Change “USP Etidronate Disodium Related Compound A RS  Sodium phosphite dibasic pentahydrate.  <math>Na_2HPO_3 \cdot 5H_2O</math> 216.04”  to:  USP Etidronate Disodium Related Compound A RS  Sodium phosphite dibasic pentahydrate.  <math>Na_2HPO_3 \cdot 5H_2O</math> 216.04 [CAS-13708-85-5]</p>
2789	<i>Etoposide Capsules</i>	<i>USP Reference standards</i> (11)	<p>Change: “USP Etoposide Related Compound A RS”  to:  USP Etoposide Resolution Mixture RS</p>
2790	<i>Etoposide Injection</i>	<i>USP Reference standards</i> (11)	<p>Change: “USP Etoposide Related Compound A RS”  to:  USP Etoposide Resolution Mixture RS</p>
2890	<i>Flurazepam Hydrochloride</i>	<i>USP Reference standards</i> (11)	<p>Change “USP Fluphenazine Enanthate RS”  to:  USP Flurazepam Hydrochloride RS</p>
2891	<i>Flurazepam Hydrochloride Capsules</i>	<i>USP Reference standards</i> (11)	<p>Change “USP Fluphenazine Enanthate RS”  to:  USP Fluphenazine Hydrochloride RS</p>
3275	<i>Letrozole Tablets</i>	<i>Assay</i>	<p>Line 1 under <i>Sample solution</i>: Change “10 <math>\mu</math>g/mL of letrozole in <i>Diluent</i>, from <i>Sample stock solution</i>”  to:  10 <math>\mu</math>g/mL of letrozole in <i>Mobile phase</i>, from <i>Sample stock solution</i></p>
3581	<i>Mycophenolate Mofetil Tablets</i>	IMPURITIES <i>Procedure 3: Limit of Z-Mycophenolate Mofetil</i>	<p>Line 3 under <i>System Suitability</i>: Change “[NOTE—The relative retention times for mycophenolate mofetil and mycophenolate Z-mycophenolate mofetil are 1.0 and 1.1, respectively.]”  to:  [NOTE—The relative retention times for mycophenolate mofetil and Z-mycophenolate mofetil are 1.0 and 1.1, respectively.]</p>
3656	<i>Nifedipine</i>	<i>USP Reference standards</i> (11)	<p>Line 11: Add “[NOTE—Nifedipine, when exposed to daylight and certain wavelengths of artificial light, readily converts to a nitrophenylpyridine derivative. Exposure to UV light leads to the formation of a nitrophenylpyridine derivative. Perform assays and tests in the dark or under golden fluorescent or other low-actinic light. Use low-actinic glassware.]”</p>
3658	<i>Nifedipine Capsules</i>	<i>USP Reference standards</i> (11)	<p>Line 11: Add “[NOTE—Nifedipine, when exposed to daylight and certain wavelengths of artificial light, readily converts to a nitrophenylpyridine derivative. Exposure to UV light leads to the formation of a nitrophenylpyridine derivative. Perform assays and tests in the dark or under golden fluorescent or other low-actinic light. Use low-actinic glassware.]”</p>

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3698	<i>Nystatin Oral Suspension</i>	<i>Uniformity of dosage units (905)</i>	Line 1 under <i>Procedure for content uniformity</i> : Change “[NOTE—Use low-actinic glassware. The correction factor, <i>F</i> , calculated as directed in section (4) of <i>Content Uniformity</i> under <i>Uniformity of Dosage Units (905)</i> , is invalid if the value obtained by the formula in the second sentence is greater than 25; follow sections (5) and (6), except to substitute 0.750 for 0.900.]” to: [NOTE—Use low-actinic glassware.]
3749	<i>Oxaliplatin for Injection</i>	IMPURITIES <i>Procedure 3: Limit of Related Compound C and Unspecified Impurities</i>	Line 7 under <i>Procedure 3</i> : Change “ <i>Mobile phase and Chromatographic system</i> .” to: <i>Mobile phase</i> :
			Line 25 under <i>Procedure 3</i> : Add “ <i>Chromatographic system</i> : Proceed as directed in the <i>Assay</i> , except for the <i>Injection size</i> . <i>Injection size</i> : 10 µL”
3758	<i>Oxazepam Tablets</i>	<i>Assay</i>	Line 1: Change “Weigh and finely powder not less than 20 Tablets. Transfer an accurately weighed portion of the powder, equivalent to about 50 mg of oxazepam, to a medium-porosity, sintered-glass funnel that is fitted into a small suction flask, and proceed as directed in the <i>Assay</i> under <i>Oxazepam Capsules</i> , beginning with “Add 25 mL of alcohol.”” to: Weigh and finely powder not less than 20 Tablets. Transfer an accurately weighed portion of the powder, equivalent to about 50 mg of oxazepam, to a medium-porosity, sintered-glass funnel that is fitted into a small suction flask. Add 25 mL of alcohol, mix with the aid of a stirring rod, and after about 5 minutes apply gentle suction to remove the extract. Repeat the extraction with four additional 25-mL portions of alcohol, transfer the extracts to a 250-mL volumetric flask, dilute with alcohol to volume, and mix. Transfer 2.0 mL of this solution to a 100-mL volumetric flask, dilute with alcohol to volume, and mix. Concomitantly determine the absorbances of this solution and of a Standard solution of USP Oxazepam RS in the same medium having a known concentration of about 4 µg per mL in 1-cm cells at the wavelength of maximum absorbance at about 229 nm, with a suitable spectrophotometer, using alcohol as the blank.
3808	<i>Pancuronium Bromide</i>	<i>USP Reference standards (11)</i>	Line 2: Delete 3α, 17β-dihydroxy-2β, 16β-dipiperidinyl-5α-androstane, 3,17-diacetate, dimethobromide. <chem>C35H60Br2N2O4</chem> 732.67
4168	<i>Risedronate Sodium Tablets</i>	<i>Assay</i>	Line 6 under <i>Assay preparation</i> : Change “0.5–1.5 g per mL” to: 0.5–1.5 mg per mL
4226	<i>Simethicone Emulsion</i>	IDENTIFICATION <i>Infrared Absorption (1975)</i>	Line 4 under <i>Analysis</i> : Delete “Place about 5 drops of the <i>Sample solution</i> in the sample trough, and dry it with a stream of nitrogen.”
4436	<i>Ticlopidine Hydrochloride</i>	SPECIFIC TESTS <i>Limit of Formaldehyde</i>	Line 1 under <i>Sample solution</i> : Change “50 mg/mL of Ticlopidine Hydrochloride in methanol” to: 0.50 g of Ticlopidine Hydrochloride in 10 mL methanol
4477	<i>Trandolapril</i>	<i>Related compounds</i>	Line 7 under <i>Chromatographic system</i> : Change “Chromatograph the <i>Resolution solution</i> , and record the responses as directed for <i>Procedure</i> : the resolution, <i>R</i> , between the peaks due to trandolapril related compound C and trandolapril related compound D is not less than 4; the tailing factor is ≤1.5; and the relative standard deviation for replicate injections is not more than 3.0%.” to: Chromatograph the <i>Resolution solution</i> , and record the responses as directed for <i>Procedure</i> : the resolution, <i>R</i> , between the peaks due to trandolapril related compound C and trandolapril related compound D is not less than 4. Chromatograph the <i>Standard solution</i> , and record the responses as directed for <i>Procedure</i> : the tailing factor is 1.5; and the relative standard deviation for replicate injections is not more than 3.0%.

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4564	Vancomycin	Chromatographic purity	<p>Change: "Triethylamine buffer, Solution A, Solution B, Mobile phase, Resolution solution, and Chromatographic system—Prepare as directed in the test for Chromatographic purity under Vancomycin Hydrochloride</p> <p><i>Test preparation A</i>—Transfer about 250 mg of Vancomycin to a 25-mL volumetric flask, add 5 mL of <i>Solution A</i>, then add 0.1 N hydrochloric acid dropwise with swirling until dissolution is achieved. Dilute with <i>Solution A</i> to volume, and mix.</p> <p><i>Test preparation B</i>—Transfer 2.0 mL of <i>Test preparation A</i> to a 50-mL volumetric flask, dilute with <i>Solution A</i> to volume, and mix.</p> <p><i>Procedure</i>—Proceed as directed for <i>Procedure</i> in the test for Chromatographic purity under Vancomycin Hydrochloride. Calculate the percentage of vancomycin B in the specimen taken by the formula: <math>2500r_B / (25r_B + r_A)</math> in which the terms are as defined therein: not less than 92% of vancomycin B is found.</p> <p>Calculate the percentage of any individual peak, other than the main peak, by the formula: <math>100r_{Ai} / (25r_B + r_A)</math> in which the terms are as defined therein: not more than 3% of any peak other than the main peak is found."</p> <p>to:</p> <p><i>Triethylamine buffer</i>—Mix 4 mL of triethylamine and 2000 mL of water, and adjust with phosphoric acid to a pH of 3.2.</p> <p><i>Solution A</i>—Prepare a mixture of <i>Triethylamine buffer</i>, acetonitrile, and tetrahydrofuran (92 : 7 : 1), and degas briefly.</p> <p><i>Solution B</i>—Prepare a suitable mixture of <i>Triethylamine buffer</i>, acetonitrile, and tetrahydrofuran (70 : 29 : 1), and degas briefly.</p> <p><i>Mobile phase</i>—Use variable mixtures of <i>Solution A</i> and <i>Solution B</i> as directed for <i>Chromatographic system</i>. Make adjustments if necessary (see <i>System Suitability</i> under <i>Chromatography</i> (621)), changing the acetonitrile proportion in <i>Solution A</i> to obtain a retention time of 7.5 to 10.5 minutes for the main vancomycin peak.</p> <p><i>Resolution solution</i>—Prepare a solution of USP Vancomycin Hydrochloride RS in water containing 0.5 mg per mL, heat at 65° for 48 hours, and allow to cool.</p> <p><i>Test preparation A</i>—Transfer about 250 mg of Vancomycin to a 25-mL volumetric flask, add 5 mL of <i>Solution A</i>, then add 0.1 N hydrochloric acid dropwise with swirling until dissolution is achieved. Dilute with <i>Solution A</i> to volume, and mix.</p> <p><i>Test preparation B</i>—Transfer 2.0 mL of <i>Test preparation A</i> to a 50-mL volumetric flask, dilute with <i>Solution A</i> to volume, and mix.</p> <p><i>Chromatographic system</i> (see <i>Chromatography</i> (621))—The liquid chromatograph is equipped with a 280-nm detector and a 4.6-mm × 25-cm column that contains 5-μm packing L1. The flow rate is about 2 mL per minute. The chromatograph is programmed as follows.</p> <table border="1" data-bbox="860 1396 1299 1606"> <thead> <tr> <th>Time (minutes)</th> <th><i>Solution A</i> (%)</th> <th><i>Solution B</i> (%)</th> <th>Elution</th> </tr> </thead> <tbody> <tr> <td>0-12</td> <td>100</td> <td>0</td> <td>isocratic</td> </tr> <tr> <td>12-20</td> <td>100—0</td> <td>0—100</td> <td>linear gradient</td> </tr> <tr> <td>20-22</td> <td>0</td> <td>100</td> <td>isocratic</td> </tr> <tr> <td>22-23</td> <td>0—100</td> <td>100—0</td> <td>linear gradient</td> </tr> <tr> <td>23-30</td> <td>100</td> <td>0</td> <td>isocratic</td> </tr> </tbody> </table> <p>Chromatograph the <i>Resolution solution</i>, and record the peak responses as directed for <i>Procedure</i>: the elution order is resolution compound 1, vancomycin B, and resolution compound 2. Resolution compound 2 elutes 3 and 6 minutes after the start of the period when the percentage of <i>Solution B</i> is increasing from 0% to 100%. The resolution, <i>R</i>, between resolution compound 1 and vancomycin B is not less than 3.0; and the column efficiency, calculated from the vancomycin B peak, is not less than 1500 theoretical plates.</p> <p><i>Procedure</i>—[NOTE—Where baseline separation is not achieved, peak areas are defined by vertical lines extended from the valleys between peaks to the baseline. The main component peak may include a fronting shoulder, which is attributed to monodechlorovancomycin. This shoulder should not be integrated separately.] Separately inject equal volumes (about 20 μL) of <i>Test preparation A</i></p>	Time (minutes)	<i>Solution A</i> (%)	<i>Solution B</i> (%)	Elution	0-12	100	0	isocratic	12-20	100—0	0—100	linear gradient	20-22	0	100	isocratic	22-23	0—100	100—0	linear gradient	23-30	100	0	isocratic
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4564	Vancomycin	Chromatographic purity, continued	<p>and <i>Test preparation B</i> into the chromatograph, record the chromatograms, and measure the area responses for all of the peaks. [NOTE—Correct any peak observed in the chromatograms obtained from <i>Test preparation A</i> and <i>Test preparation B</i> by subtracting the area response of any peak observed in the chromatogram of Solution A at the corresponding elution time.] Calculate the percentage of vancomycin B in the specimen tested by the formula: <math>2500r_B / (25r_B + r_A)</math> in which <math>r_B</math> is the corrected area response of the main peak obtained in the chromatogram of <i>Test preparation B</i>; and <math>r_A</math> is the sum of the corrected area responses of all the peaks, other than the main peak, in the chromatogram obtained from <i>Test preparation A</i>: not less than 92% of vancomycin B is found. Calculate the percentage of each other peak taken by the formula: <math>100r_{Ai} / (25r_B + r_A)</math> in which <math>r_{Ai}</math> is the corrected area response of any individual peak, other than the main peak, obtained in the chromatogram of <i>Test preparation A</i>: not more than 3% of any peak other than the main peak is found.</p>
4567	Vancomycin Injection	Chromatographic purity	<p>Change: "<i>Triethylamine buffer, Solution A, Solution B, Mobile phase, and Chromatographic system</i>—Prepare as directed in the test for <i>Chromatographic purity</i> under <i>Vancomycin Hydrochloride</i>. <i>Resolution solution</i>—Allow a container of Injection to thaw, and mix the solution. Dilute a portion of the solution with water to obtain a solution containing 0.5 mg of vancomycin per mL, heat at 65° for 24 hours, and allow to cool. <i>Test preparation A</i>—Allow a container of Injection to thaw, and mix the solution. <i>Test preparation B</i>—Transfer 2.0 mL of <i>Test preparation A</i> to a 50-mL volumetric flask, dilute with <i>Solution A</i> to volume, and mix. <i>Procedure</i>—Proceed as directed for <i>Procedure</i> in the test for <i>Chromatographic purity</i> under <i>Vancomycin Hydrochloride</i>. Calculate the percentage of vancomycin B in the specimen taken by the formula: <math>2500r_B / (25r_B + r_A)</math> in which the terms are as defined therein: not less than 88% of vancomycin B is found. Calculate the percentage of any individual peak, other than the main peak, by the formula: <math>100r_{Ai} / (25r_B + r_A)</math> in which the terms are as defined therein: not more than 4% of any peak other than the main peak is found." to: <i>Triethylamine buffer</i>—Mix 4 mL of triethylamine and 2000 mL of water, and adjust with phosphoric acid to a pH of 3.2. <i>Solution A</i>—Prepare a mixture of <i>Triethylamine buffer</i>, acetonitrile, and tetrahydrofuran (92 : 7 : 1), and degas briefly. <i>Solution B</i>—Prepare a suitable mixture of <i>Triethylamine buffer</i>, acetonitrile, and tetrahydrofuran (70 : 29 : 1), and degas briefly. <i>Mobile phase</i>—Use variable mixtures of <i>Solution A</i> and <i>Solution B</i> as directed for <i>Chromatographic system</i>. Make adjustments if necessary (see <i>System Suitability</i> under <i>Chromatography</i> (621)), changing the acetonitrile proportion in <i>Solution A</i> to obtain a retention time of 7.5 to 10.5 minutes for the main vancomycin peak. <i>Resolution solution</i>—Allow a container of Injection to thaw, and mix the solution. Dilute a portion of the solution with water to obtain a solution containing 0.5 mg of vancomycin per mL, heat at 65° for 24 hours, and allow to cool. <i>Test preparation A</i>—Allow a container of Injection to thaw, and mix the solution. <i>Test preparation B</i>—Transfer 2.0 mL of <i>Test preparation A</i> to a 50-mL volumetric flask, dilute with <i>Solution A</i> to volume, and mix. <i>Chromatographic system</i> (see <i>Chromatography</i> (621))—The liquid chromatograph is equipped with a 280-nm detector and a 4.6-mm × 25-cm column that contains 5-µm packing L1. The flow rate is about 2 mL per minute. The chromatograph is programmed as follows.</p>

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4567	Vancomycin Injection	Chromatographic purity, continued	<table border="1"> <thead> <tr> <th>Time (minutes)</th> <th>Solution A (%)</th> <th>Solution B (%)</th> <th>Elution</th> </tr> </thead> <tbody> <tr> <td>0-12</td> <td>100</td> <td>0</td> <td>isocratic</td> </tr> <tr> <td>12-20</td> <td>100→0</td> <td>0→100</td> <td>linear gradient</td> </tr> <tr> <td>20-22</td> <td>0</td> <td>100</td> <td>isocratic</td> </tr> <tr> <td>22-23</td> <td>0→100</td> <td>100→0</td> <td>linear gradient</td> </tr> <tr> <td>23-30</td> <td>100</td> <td>0</td> <td>isocratic</td> </tr> </tbody> </table> <p>Chromatograph the <i>Resolution solution</i>, and record the peak responses as directed for <i>Procedure</i>: the elution order is resolution compound 1, vancomycin B, and resolution compound 2. Resolution compound 2 elutes 3 and 6 minutes after the start of the period when the percentage of <i>Solution B</i> is increasing from 0% to 100%. The resolution, <i>R</i>, between resolution compound 1 and vancomycin B is not less than 3.0; and the column efficiency, calculated from the vancomycin B peak, is not less than 1500 theoretical plates.</p> <p><i>Procedure</i>—[NOTE—Where baseline separation is not achieved, peak areas are defined by vertical lines extended from the valleys between peaks to the baseline. The main component peak may include a fronting shoulder, which is attributed to monodechloro-vancomycin. This shoulder should not be integrated separately.] Separately inject equal volumes (about 20 µL) of <i>Test preparation A</i> and <i>Test preparation B</i> into the chromatograph, record the chromatograms, and measure the area responses for all of the peaks. [NOTE—Correct any peak observed in the chromatograms obtained from <i>Test preparation A</i> and <i>Test preparation B</i> by subtracting the area response of any peak observed in the chromatogram of <i>Solution A</i> at the corresponding elution time.] Calculate the percentage of vancomycin B in the specimen tested by the formula:</p> $2500r_B / (25r_B + r_A)$ <p>in which <math>r_B</math> is the corrected area response of the main peak obtained in the chromatogram of <i>Test preparation B</i>; and <math>r_A</math> is the sum of the corrected area responses of all the peaks, other than the main peak, in the chromatogram obtained from <i>Test preparation A</i>: not less than 88% of vancomycin B is found. Calculate the percentage of each other peak taken by the formula:</p> $100r_{Ai} / (25r_B + r_A)$ <p>in which <math>r_{Ai}</math> is the corrected area response of any individual peak, other than the main peak, obtained in the chromatogram of <i>Test preparation A</i>: not more than 4% of any peak other than the main peak is found.</p>	Time (minutes)	Solution A (%)	Solution B (%)	Elution	0-12	100	0	isocratic	12-20	100→0	0→100	linear gradient	20-22	0	100	isocratic	22-23	0→100	100→0	linear gradient	23-30	100	0	isocratic
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4596	Water for Injection	ADDITIONAL REQUIREMENTS	Line 1: Change “[NOTE—Required for bulk and packaged forms of <i>Water for Injection</i> .]” to: [NOTE—Required for packaged forms of <i>Water for Injection</i> .]																								
4598	Purified Water	MONOGRAPH TITLE	Line 1: Change “[NOTE—Required for bulk and packaged forms of <i>Purified Water</i> ]” to: [NOTE—For microbiological guidance, see general information chapter <i>Water for Pharmaceutical Purposes</i> (1231).]																								
		DEFINITION	Line 1: Delete “[NOTE—For microbiological guidance, see general information chapter <i>Water for Pharmaceutical Purposes</i> (1231).]”																								
		SPECIFIC TESTS	Line 1: Add “[NOTE—Required for bulk and packaged forms of <i>Purified Water</i> ]”																								
4623	Zinc Carbonate	Insoluble matter	Line 5: Change “20 mg” to: 2 mg																								
4636	Zolpidem Tartrate Extended-Release Tablets	USP Reference standards (11)	Change “USP Zonisamide Related Compound A RS 1,2-Benzisoxazole-3-methanesulfonic acid sodium salt. C <sub>8</sub> H <sub>6</sub> NNaO <sub>4</sub> S 235.19 [CAS-73101-64-1]” to: USP Zolpidem Related Compound A RS <i>N,N</i> -Dimethyl-2-(7methyl-2-p-tolylimidazo[1,2- <i>a</i> ]pyridin-3-yl)acetamide. C <sub>19</sub> H <sub>21</sub> N <sub>3</sub> O 307.39																								

Page Number	Title	Section	Description
<i>First Supplement to USP 34–NF 29</i>			
5034	<i>Tamsulosin Hydrochloride Capsules</i>	PERFORMANCE TESTS <i>Dissolution, Test 8</i>	Line 1 under <i>Apparatus 2</i> : Change “50 rpm, with sinkers” to: 100 rpm, with sinkers